

**Lecture 8:****Cell-Surface Interactions: Host Responses to Biomaterials (Part II)**

Implantation of a biomaterial initiates the **inflammatory** response:

- response of vascularized tissue to local injury
- severity indicates biocompatibility of material

Cooperative Signaling Cascades:

- |   |   |                                |
|---|---|--------------------------------|
| <ol style="list-style-type: none"> <li>1. Coagulation Cascade</li> <li>2. Complement Alternative Pathway</li> </ol> | } | initiated by adsorbed proteins |
|---|---|--------------------------------|

The **complement** is a component of the immune system.

**Immune system function:** to protect against pathogens

**Innate (Native) Immunity**

- first line of defense
- nonspecific response to invading pathogens
- elicits adaptive response

**Adaptive (Acquired) Immunity**

- specificity to distinct foreign biomolecules (antigens)
- memory of exposure

**Physical/chemical barriers:**  
epithelia, antimicrobial proteins

**Blood proteins:** complement; cytokines (regulatory)

**Cells:** phagocytes (macrophages, neutrophils), natural killer cells

**Blood proteins:**  
antibodies (immunoglobulins), cytokines

**Cells:** lymphocytes (T cells, B cells)

## Complement

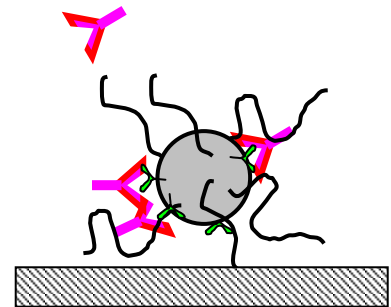
- system of >30 proteins that mediate immune response
- discriminates “foreign” from “self” through adsorbed proteins/ protein fragments (C3b, C4b)
- recruit and activate phagocytes (C3a, C5a)
- lysis of pathogens via membrane pore formation (C5b, C6-C9)

### 3 recognized pathways (to C5 convertase)

#### Classical pathway:

antigen-antibody immune complex (IC)

binds and activates C1 (autocatalytic proteolysis)  
initiating an enzymatic cascade



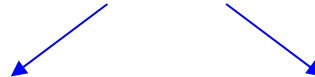
C1 → C1s

C4 → C4b

C2 → C2b

C3 → C3b

C5 → C5a/C5b



soluble fragment (16 kDa):  
recruits phagocytes by chemotaxis

insoluble fragment (170 kDa):  
initiates membrane attack complex  
(MAC) C5b•C6•C7•C8•C9

MAC pore formation compromises  
bacterial cell membrane

**Lectin pathway** (discovered in 1990's):

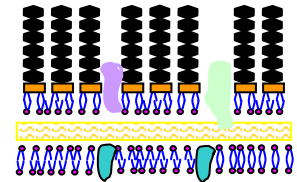
mannan binding lectin (MBL)  
binds carbohydrates on pathogen

MBL-associated serine proteases (MASP-1, -2)  
complexes with MBL

activated MASP's cleave C4 → C4b

remaining cascade follows classical pathway

bacteria cell wall  
(gram negative)

**Alternative pathway:**

nonselective pathway of complement (any foreign surface)

C3 → C3b occurs continuously in plasma at low frequency

C3b adsorbs on foreign surfaces (biomaterial)

cofactor B → C3b•Bb complex

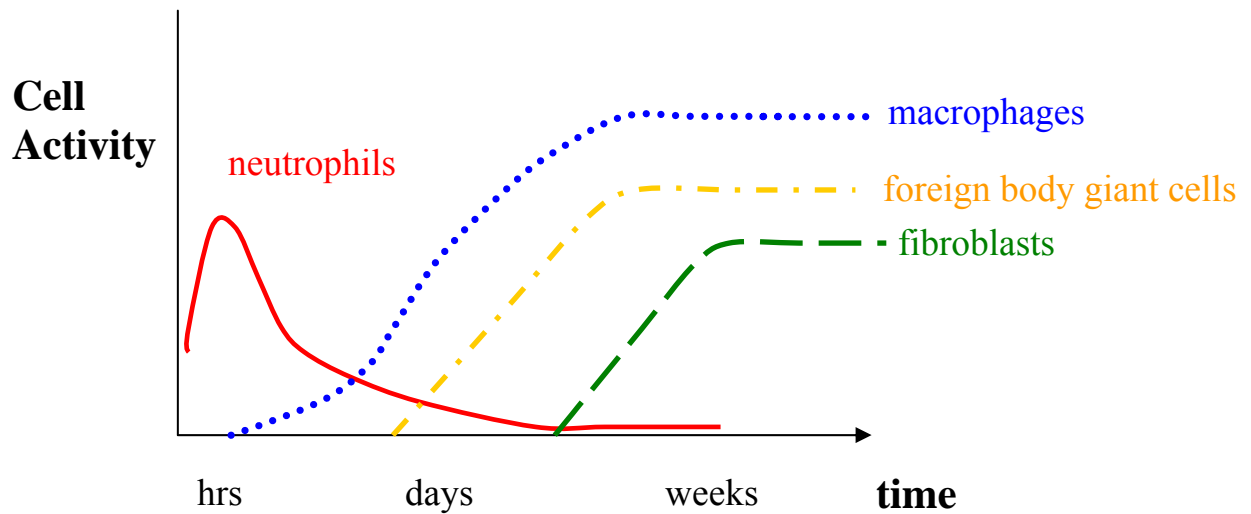
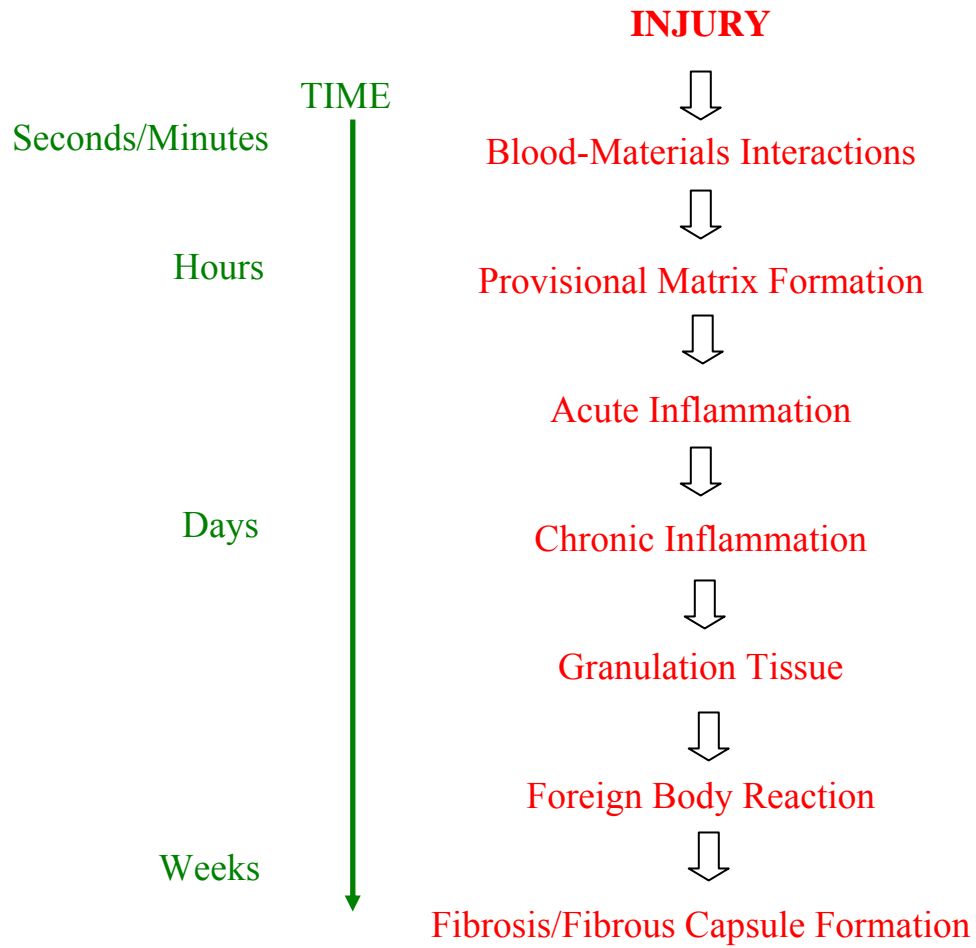
amplifies C3 → C3b

C3b•C3b•Bb complex

C5 → C5a/C5b

Soluble complement protein fragments C3a and C5a recruit phagocytes  
to site of injury

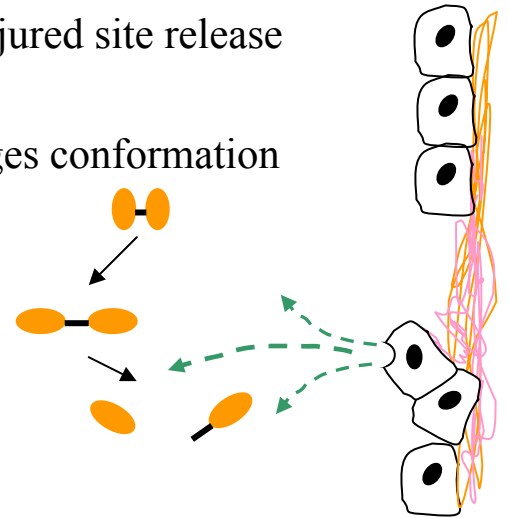
## Inflammatory Response to Implanted Biomaterials



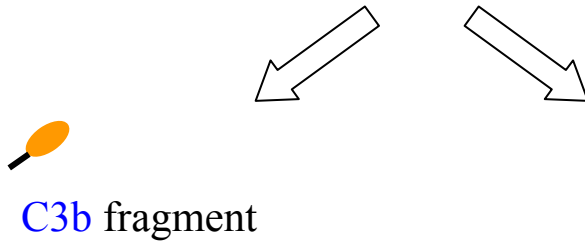
- **Endothelial cells** lining capillaries near injured site release enzymes

plasma protein **C3** (1.3 mg/ml) changes conformation

⇒ **cleaved** into fragments



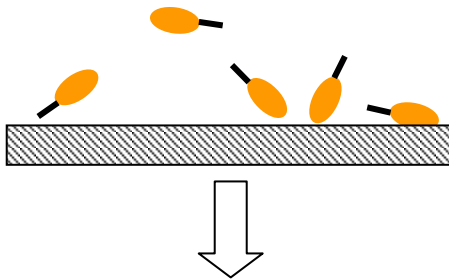
**C3**



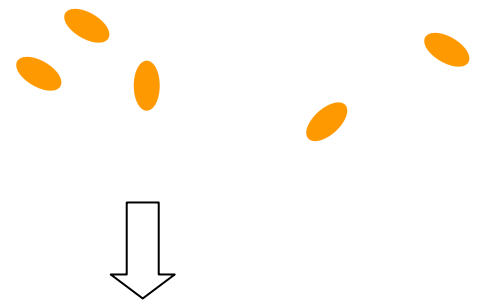
**C3b** fragment

**C3a** fragment

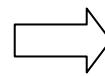
C3b attaches to biomaterial or injurious agent surface ⇒ **insoluble ligand** for **leukocyte receptors**



C3a diffuses into medium ⇒ **soluble ligand** for **leukocyte receptors**



C3b catalyzes **C5** cleavage to **C5a** ⇒ **soluble ligand** for **leukocyte receptors**

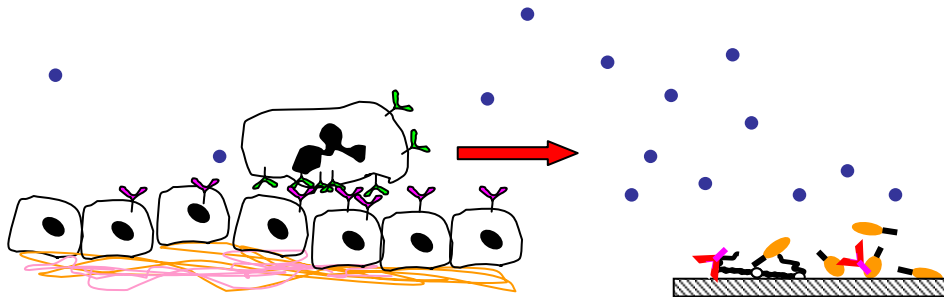


**IMMUNE CELL  
RECRUITMENT**

## Neutrophils (PMN's, polymorphonuclear leukocytes)

Associated with **acute inflammatory response** (minutes→1-2 days)

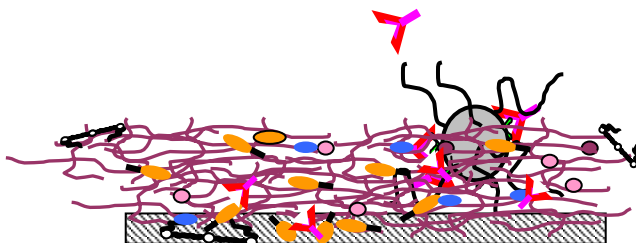
- “**first responders**” 3-5M/ml (short-lived)
- **bind C3a/C5a** via complement receptors (CR's)
- become **hyperadherent** by ↑ CR3 (integrin CD11b/CD18) surface expression – **attach to vasculature** via endothelial ICAMs
- **chemotactic to C5a**: migrate to inflammation site



On site, neutrophils **bind to C3b**, catalyzing **release of cytotoxic species**:  $H_2O_2$ ,  $O_2^- \cdot$  (superoxide radical),  $OH \cdot$ , enzymes

⇒ attack/engulf/degrade invading microbes

Released products from neutrophils, activated platelets and endothelial cells, along with fibrin, form the **provisional matrix**



- scaffold for cell attachment

- sustained release of signaling molecules

## Monocytes (0.2-0.6M/ml)

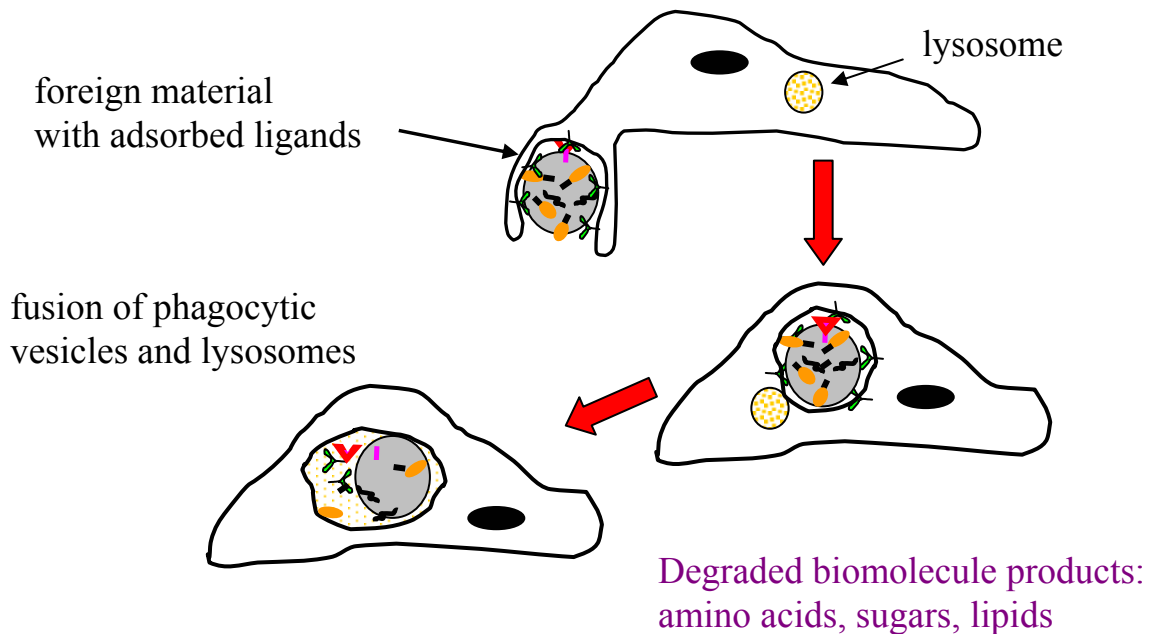
- bind C3a/C5a ⇒ follow the course of neutrophils
- Evolve to **macrophages**
- Associated with **chronic inflammation**  
days → weeks/months (or even a lifetime)



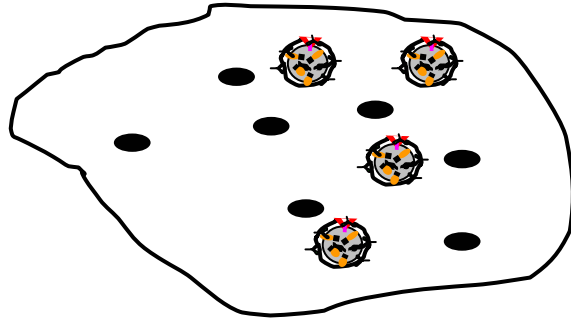
On site, macrophages **bind C3b**, secrete reactive species, enzymes, cytokines (immune cell regulators, ex. IL-1), fibronectin, growth factors (ex. fibroblast growth factor, epidermal growth factor), coagulation factors

## Macrophage response depends on foreign material properties...

- **fluids** or **small particles** (micron-sized)  
→ engulfed & degraded “**phagocytosis**”



- Numerous particulate debris or materials with high roughness
  - fusion of macrophages into multinuclear foreign body giant cells (FBGCs)



- smooth, inert implants
  - FBGCs absent (nothing to engulf)
  - macrophage layer surrounds implant

Macrophage/FBGC products (FN, FGF) recruit fibroblasts

**Fibroblasts** (connective tissue cells)

- deposit collagen
  - pink “granulation tissue” (appears in 3-5 days)
- accompanied by capillary sprouting (angiogenesis)

Wound healing histology: foreign body reaction

presence of FBGCs/macrophages, granulation tissue, capillaries at tissue/material interface

- Connective tissue remodeling ⇒ thin, encapsulating fibrous layer (fibrosis) isolates implant and foreign body rxn (weeks)



Photos removed for copyright reasons.

Fibrous capsule formation around porous P(HEMA-co-MMA) nerve conduit (J.S. Belkas et al., *Biomaterials* **26** (2005) 1741.)

FBGC formed at implant site. Arrows point to nuclei. (J.S. Belkas et al., *Biomaterials* **26** (2005) 1741.)

Formation of **scar tissue** vs. **parenchymal tissue** (tissue of specialized function) depends on:

- extent of parenchymal tissue damage (esp. tissue framework)
- parenchymal cell proliferation capacity

### Cell Regeneration Capability

<b>Category</b>	<b>Normal replic. rate</b>	<b>Response to injury</b>	<b>Examples</b>
renewing/ labile	High; via stem cell differentiation	modest ↑	skin, intestinal mucosa, bone marrow
Expanding/ stable	Low	large ↑	endothelium, fibroblasts, hepatocytes, osteoblasts
Static/ permanent	None	No replication	heart muscle cells, nerve cells

Implant **biocompatibility** is assessed largely by intensity & duration of the **inflammatory response**.

<b>Materials Class</b>	<b>Inflammatory response</b>
Metals	very severe in absence of passive oxides
Oxides	minimal
Processed natural polymers	severe
synthetic polymers	mild, unless particulate morphology; additives can give response

## **Biomaterial Biocompatibility Concerns**

### **1. Chronic inflammation**

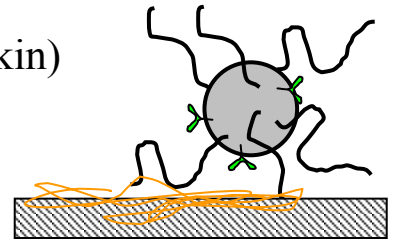
- prolonged local chemical or physical irritation—delayed healing
- often due to **moving parts**, debris, roughness
- proliferation of connective tissue, or tissue necrosis  
(2 extremes of macrophage response)

ex. PE cup liners in hip replacement implants



## 2. Bacterial Infection

- **Bacteria** compete with cells to adhere to surface
  - similar mechanisms; **better adapted to nonviable surfaces**
  - resistant to antibiotics (different surface expression)
- Most common bacterial infections:
  - **polymeric** biomaterials: *S. epidermidis* (on skin)
  - **metallic** biomaterials: *S. aureus*
  - have receptors for fibronectin & collagen



ex. artificial hearts, synthetic vessels, joint replacement implants, fixation devices, IV catheters, urologic devices, contact lenses

~60,000 U.S. deaths/yr from device-related infections  
 urinary catheters, central venous catheters

## 3. Blood Incompatibility

- blood-materials interactions lead to clot or **thrombus**
  - may compromise device by **occlusion**  
 ex. small (< 5 mm dia.) vascular grafts, stents, IV catheters
  - may detach (**embolize**) & create vessel occlusion downstream  
 ex. emboli to brain from mechanical heart valves ⇒ stroke
  - susceptible devices require use of **anti-coagulation drugs**  
 (heparin) ⇒ **bleeding risk**

- complement activation by extracorporeal therapies
  - C3b adsorption to material  $\Rightarrow$  C5a activation of neutrophils & monocytes (WBCs) to hyperadherent state
  - WBCs stick in lungs  $\Rightarrow$  neutropenia, respiratory distress, hypoxemia (O<sub>2</sub> deficiency—similar symptoms to altitude sickness), tachycardia, cardiac arrest

ex. hemodialysis membranes, cardiopulmonary bypass (CPB) devices

#### 4. Toxicity

- classical toxicity: from corrosion, degradation or wear products; cytotoxicity increases with amount present
- immune system toxicity:
  - i) immunogenic substances: proteins, carbohydrates, lipids (weakly)  
ex. processed collagen, natural latex
  - ii) small molecules (metals, degradation products, drugs) bind on host proteins/cells, making an innocuous substance antigenic  
ex. hypersensitivity to metals, acrylics



## 5. Tumorigenesis

- rarely observed
- **morphology dependent** vs. chemistry dependent  
(ex. asbestos – needle-like particulates, aspect ratio > 100:1)
- requires fibrous encapsulation (not seen at chronic inflammation sites)
- implant role unclear—foreign body reaction may stimulate maturation & proliferation of precancerous cells
- chemical carcinogens: little supportive data
  - **metal implant debris** (Cr, Co, Ni) ⇒ carcinogenic in rodents
  - **polymer impurities/additives**: monomers, solvents, plasticizers, antioxidants

### References:

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D.A. Hammer and M. Tirrell, “Biological Adhesion at Interfaces”, *Annu. Rev. Mater. Sci.* 1996, **26**: 651-691.

K.M. Yamada, “Cell Adhesion Molecules”, in *Encyclopedia of Molecular Biology*, T.E. Creighton, Ed., John Wiley & Sons: NY, 1999, pp. 361-366.

J.M. Anderson, “Biological Responses to Materials”, *Annu. Rev. Mater. Res.* 2001, **31**: 81-110.