Harvard-MIT Division of Health Sciences and Technology HST.508: Quantitative Genomics, Fall 2005 Instructors: Leonid Mirny, Robert Berwick, Alvin Kho, Isaac Kohane

Welcome to HST.508/Biophysics 170

Our emphasis

- Evolution
- Quantitative
- Medical applications

Syllabus

- 1. Evolutionary and population genetics
- 2. Comparative genomics
- 3. Structural genomics and proteomics
- 4. Functional genomics and networks

Module 1

- 1. Evolutionary and population genetics
- The basic forces of evolution: mutation, recombination, mating, migration. Neutral evolution and <u>drift</u>, effective population size, coalescent theory.
- Selection, fitness, and diffusion models. Selection at genetic and higher levels
- <u>Phylogenetic analysis.</u> Models of nucleotide evolution: <u>Jukes-Cantor</u>, Kimura, maximum likelihood models; Human/mouse/rate examples.
- **Measuring selection:** from 'classical' methods to <u>maximum</u> <u>likelihood</u> (with applications to disease evolution, HIV and influenza)
- Medical Lecture: Genetic diversity and evolution of hepatitis C virus

Drift, mutations, selection

Consider a deleterious recessive allele A_1 of frequency p in a randomly mating human population with mutations, selection and drift. Steady state distribution of p is given by

$$P(p) = C \exp(-2Nsp^2)(1-p)^{4Nu_1-1}p^{4Nu_2-1}$$
(1)

where N is an effective population size, s is selection coefficient, u_1 and u_2 are mutation rates to A₁ and from A₁ respectively, and C is a normalization coefficient.

Module 2

- 2. Comparative genomics
- <u>Sequence comparison</u>, <u>substitution matrices</u>, alignment methods, alignment statistics. <u>Multiple alignments</u>, profiles and PSSMs.
- Genome comparison and genome evolution: duplication, recombination, insertions, repeats. Orthologs, paralogs, in/outparalogs. Algorithms of genome alignment.
 Conserved non-coding, positive selection. Motif discovery.
- Prediction of gene function using: homology, context, structure, networks.
- <u>SNPs</u>: microevolution, history of population, markers medical applications (<u>example</u>)
- <u>Medical Lecture</u> Finding the keys to human heart disease in the genomes of other animals.



Figure by MIT OCW.

Model for Sequence evolution (DNA): Each site of the DNA sequence evolves according to a Markov Chain with state space {A,C,G,T}.



Figure by MIT OCW.

MARKOV CHAIN

Let $X_0, X_1, X_2, X_3, \ldots$ be a Markov chain with state space S, for example $S = \{a, c, g, t\}$.

TRANSITION MATRIX

$$P = \begin{pmatrix} p_{a,a} & p_{a,c} & p_{a,g} & p_{a,t} \\ p_{c,a} & p_{c,c} & p_{c,g} & p_{c,t} \\ p_{g,a} & p_{g,c} & p_{g,g} & p_{g,t} \\ p_{t,a} & p_{t,c} & p_{t,g} & p_{t,t} \end{pmatrix}$$

Here

$$p_{i,j} = \mathbf{P}(X_{n+1} = j | X_n = i)$$

for $n \ge 0$, where $i, j \in \{a, c, g, t\}$.

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Simplest model for sequence evolution: Jukes-Cantor

$$\begin{pmatrix} p_{a,a} & p_{a,c} & p_{a,g} & p_{a,t} \\ p_{c,a} & p_{c,c} & p_{c,g} & p_{c,t} \\ p_{g,a} & p_{g,c} & p_{g,g} & p_{g,t} \\ p_{t,a} & p_{t,c} & p_{t,g} & p_{t,t} \end{pmatrix} = \begin{pmatrix} 1-3\alpha & \alpha & \alpha & \alpha \\ \alpha & 1-3\alpha & \alpha & \alpha \\ \alpha & \alpha & 1-3\alpha & \alpha \\ \alpha & \alpha & \alpha & 1-3\alpha \end{pmatrix}$$

The stationary distribution is $\vec{\pi} = (0.25, 0.25, 0.25, 0.25).$

The parameter α depends on the time scale

(if the unit time is 100.000 generations, α would take a smaller value than if the unit time were chosen as 200.000 generations).

Necessary: $\alpha < 1/3$.

The *n*-step transition probabilities can be computed: $\mathbf{P}(X_n = i | X_0 = i) = 0.25 + 0.75 \cdot (1 - 4\alpha)^n$, for $i \in \{a, c, g, t\}$. $\mathbf{P}(X_n = j | X_0 = i) = 0.25 - 0.25 \cdot (1 - 4\alpha)^n$, for $i, j \in \{a, c, g, t\}, i \neq j$. Underlying Model: Each site in the sequence evolves according to a Markov chain, and independently of the other sites.



Figure by MIT OCW.

FROM TRANSITION MATRIX TO ALIGNMENT SCORES

Two hypothesis:

- 1. Sequences S1 and S2 are unrelated (=random matching)
- 2. Sequences S1 and S2 have a common ancestor.

Score = Log (P1/P2)

P1 - probability of observed alignment given model 1P2 - probability of observed alignment given model 2

Homology



Figure by MIT OCW.

Guest lecture 1

- Richard Lewontin
 - evolutionary geneticist
 - philosopher of science
 - social critic
 - numerous publications including, "The Spandrels of San Marco", "The Genetic Basis of Evolutionary Change", "Biology as Ideology", "The Triple Helix: Gene, Organism, and Environment" ...
 - http://hrst.mit.edu/hrs/evolution/public/profiles/lewontin.html
 - http://www.nybooks.com/authors/4463

(Compilation of information by MIT OCW.)

Module 3

- 3. Structural genomics and proteomics
- Overview of <u>protein structures</u>, <u>domain architecture</u>. Sequence-structure mapping, protein folding, forces and interactions.
- Structure-based substitution matrices. Protein structure prediction. Threading.
- Protein function: binding and kinetics. Michaelis-Menthen kinetics, inhibition. Protein-DNA recognition: models and algorithms.
- Proteomics: networks of protein-protein interactions, complexes, modules. Power-law distributions, clustering coefficient. Evolution of networks.
- <u>Medical Lecture Hemoglobin and the anemias.</u>

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ELECTRO + SOLVENT : Dielectric effect $V = \frac{q_i q_j}{4\pi \epsilon r_{ij}}; \quad \epsilon = 80$

Figure removed due to copyright considerations.

2-3 Kcal/mol ∆G ~ T

Linear in T => entropic!

More forces:



Hydrophobic effect

Frank & Evans 1945

- Water molecules form hydrogen bonds
- Polar groups do not disturb the network of water-water interactions.
- Non-polar (hydrophobic) groups disrupt the network leading to formation of "local ordering" of water.
- Local ordering reduces the entropy

Figure removed due to copyright reasons.

Please see Figure 2 in:

Laidig, Keith E., and Valerie Daggett. "Testing the Modified Hydration-Shell Hydrogen-Bond Model of Hydrophobic Effects Using Molecular Dynamics Simulation." *J Phys Chem* 100 (1996): 5616-5619.

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RECOGNITION OF BINDING MOTIFS IN DNA

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RECOGNITION OF BINDING MOTIFS IN DNA 2

HOMEODOMAIN

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Examples of biological networks

 Protein-protein interactions proteins ~5000 interactions ~7000

Figures removed due to copyright reasons.

Databases: BindDB MIPS DIP

A comprehensive analysis of protein-protein interactions in S.cerevisiae. *Nature.* 2000, 623-7. Uetz P et al

Examples of biological networks

Protein-DNA interactions (TF-upstream binding)

Figures removed due to copyright reasons.

Please see Figure 5 in T. I., Lee, et al. "Transcriptional regulatory networks in Saccharomyces cerevisiae." *Science* 298, no. 5594 (Oct 25, 2002): 799-804.

Metabolic Pathways

Figure removed due to copyright considerations.

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it's not a random graph!

Figures removed due to copyright reasons. Please see figures 1e and 2 in Jeong, H., B. Tombor, R. Albert, Z. N. Oltvai, and A. L. Barabasi. "The large-scale organization of metabolic networks." *Nature* 407, no. 6804 (Oct 5, 2000): 651-4.

IT'S ALMOST SCALE-FREE (=POWER-LAW) GRAPH

Figures removed due to copyright reasons. Please see:

Liljeros, F., et al. "Distributions of number of sexual partnerships have power law decaying tails and finite variance." eprint ARXIV, (http://arxiv.org/). (May 2003).

and

Liljeros F., et al. "The web of human sexual contacts." Nature 411, no. 6840 (Jun 21, 2001): 907-8.

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Module 4

4. Functional Genomics and Networks

- Gene regulation and function, conservation, detecting regulatory elements.
- RNA expression: clustering and classification.
- RNA expression: classification, 2-way clustering, regulatory modules. Integration of expression and proteomic data.
- **Dynamics of biological networks metabolic, regulatory.** FBA, signaling, regulation of gene expression.
- <u>Medical Lecture:</u> Two examples: phenylketonuria (monogenic) and diabetes type 2 (multigenic+). "Disease" genes vs. "susceptibility" genes. "Environmental" vs. "Developmental" regulation of gene expression.

cDNA microarray expt



Figure by MIT OCW.



Figure by MIT OCW.

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DEFINITION OF THE CLUSTERING PROBLEM



CLUSTER ANALYSIS YIELDS DENDROGRAM

T (RESOLUTION)



BUT WHAT ABOUT THE OKAPI?



Figure by MIT OCW.



how many clusters?

3 LARGE MANY small (SPC)

Figures derived from Blatt, M., S. Wiseman, and E. Domany. "Superparamagnetic Clustering of Data." *Phys Rev Lett* 76, no. 18 (1996): 3251.

OTHER METHODS



Figures derived from Blatt, M., S. Wiseman, and E. Domany. "Superparamagnetic Clustering of Data." *Phys Rev Lett* 76, no. 18 (1996): 3251.

Functionally related genes

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Figure by MIT OCW.

HST.508 Cases

Case 1: Dr. Gail Genomous

- Took HST.508 in 2005
- Since 2008 works at R&D of Shrek Pharmaceuticals
- She works on target identification for mysterious green-rash syndrome.
 - Pathways involved are know. But candidate drugs although binding to their targets have been inefficient on model organisms.
 - Her goals are
 - Suggest better target proteins
 - Assist drug design/organic syntheses group in drug development

Case 1: Dr. Gail Genomous

- She found that involved drug targets are enzymes of wellknown metabolic pathway. She sets up flux-balance simulations to study this pathways (Lec F4).
- Simulations suggest that inhibition of targeted enzymes does not shutoff the pathway. Dr.Genomous identifies other enzymes that need to be inhibited to shutoff the pathway. She suggests that down-regulating these enzymes is the most efficient intervention strategy.

Case 1: Dr. Gail Genomous

- She finds out that these enzymes are co-expressed (Lec F1), but transcription factor is unknown.
- However, available protein-protein interactions suggest that these enzymes are also activated by a kinase pi314 (Lec S4).
- Dr.Genomous model the structure of pi314 by homology to another kinase (Lec S1).
- Comparison of pi314 with its orthologs from related species (*P.Winnie*, *D.Scooby* etc) suggests functional region of the structure (Lec C2,S2).
- Dr. Genomous proposes drug design group the new drug target and a specific functional region to be targeted.

Case 2: Dr. Pete Proteomson

- Took HST.508 in 2005
- Works on his dissertation at the Whitetail institute
- He is interested in tail discoloration syndrome (TDS).
 - In collaboration with the hospital, his lab has performed expression profiling of patients and normal individuals.
 - His goals are
 - Identify genes involved
 - Suggest and test mechanisms of the disease

Case 2: Dr. Pete Proteomson

- Dr Proteomson has detected differentially expressed genes (Lecture F2)
- Mapped differentially expressed genes on the network of protein-protein interactions (Lec. S3) and network of synthetically lethal genes. Results suggested that most of these genes belong to the same pathway (Lec S4).
- Using homology, genomic and network location, he predicted function for some of these genes as protein kinases involved in cell signaling (Lec S4).

Case 2: Dr. Pete Proteomson

- Mapping selected gene onto a database of SNPs revealed rear polymorphisms in some of these genes (Lec C4).
- Population analysis suggests that these mutations are likely to be deleterious (Lec E3). Mapping mutations on known structures of protein kinases and cross-species comparison strongly supported deleterious nature of some SNPs (Lec S2,C4).
- Finally, Dr.Proteomson demonstrated that patients carry more deleterious SNPs in the identified pathway than normal individuals. This result supports the role of identified pathways in development of the green rash.