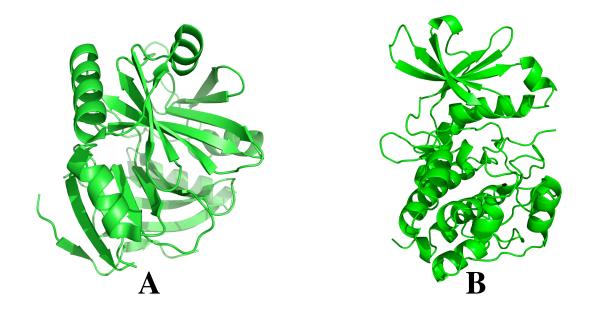
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5.36 Biochemistry Laboratory Spring 2009

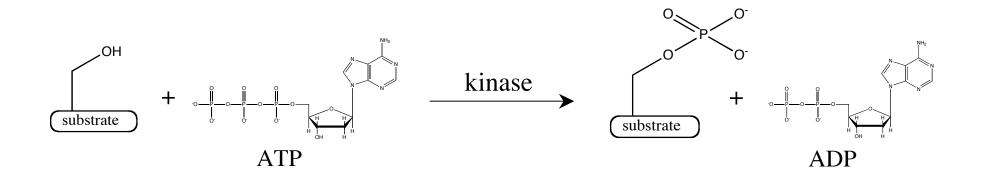
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Which is a kinase domain? A, B, or both?

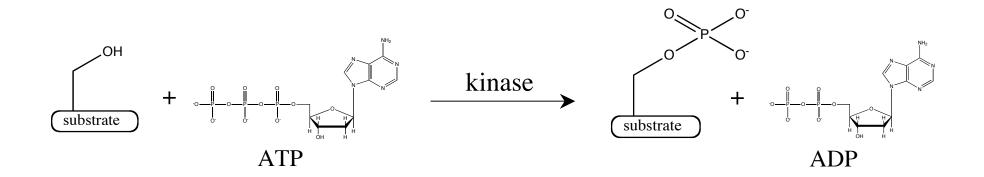
Detecting Kinase Activity in vitro and in vivo

- I. In vitro activity assays (for purified kinases)
 - A. ³²P incorporation assay
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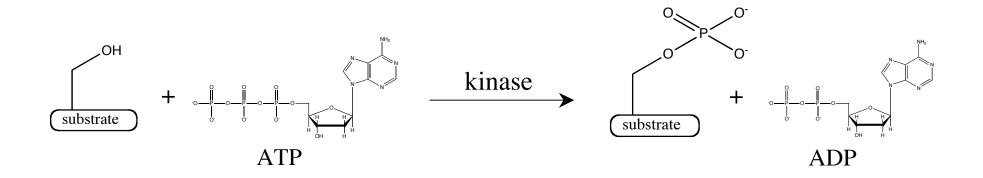


Why are scientists interested in detecting kinase activity?

- Enables monitoring of kinase activity and inhibition
- Used for screening of drug candidates
- Facilitates the unraveling of complex cell-signalling pathways implicated in processes ranging from cell cycle regulation to cellular migration (including tumor metastasis).



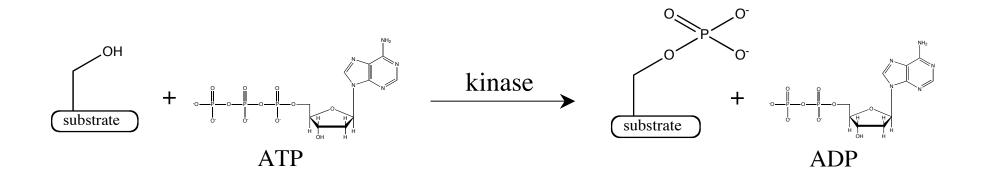
Enzyme activity assays detect either product formation or starting material consumption.



Enzyme activity assays detect either product formation or starting material consumption.

For kinases this means

- Detection of phosphorylated substrate (product) or
- Detection of ADP (product) formation or ATP (starting material) consumption



However, phosphorylation is spectroscopically silent.

Neither the formation of phosphorylated product nor the conversion of ATP to ADP can be monitored *directly* using spectroscopy.

The products appear identical to the starting materials due to identical absorption features and extinction coefficients.

Kinase assays: in buffer, in cell lysate, in vivo

Depending on your application, you should consider whether you need an assay that can be performed only with purified enzyme, in a cell lysate mixture, or in vivo.

Some other considerations may include a protocol that is:

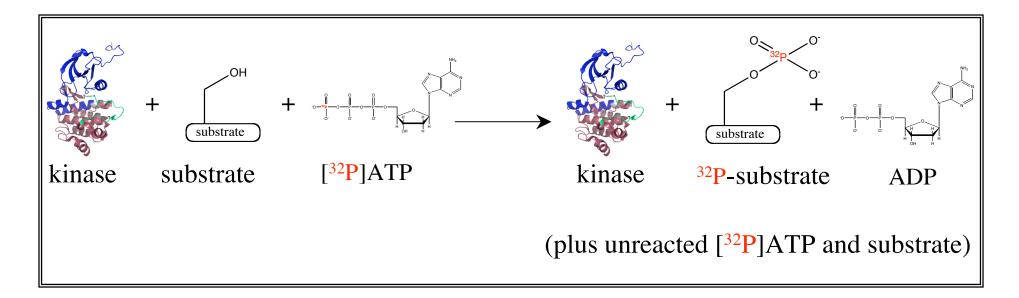
- Easy and safe to use
- General versus specific
- Continuous (versus an end-point assay)
- A turn-on versus a turn-off assay (some researchers prefer the generation of a positive signal rather than a decrease in signal)
- Amenable to high-throughput applications
- Sensitive to low levels of kinase activity

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³²P incorporation assay

Detection of a radiolabeled phosphorylated substrate

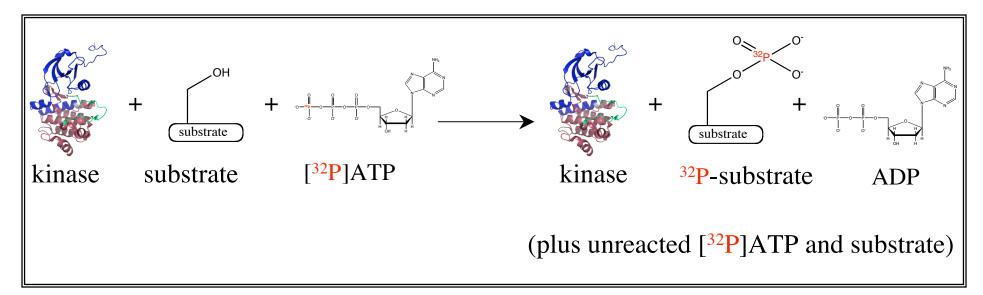


The kinase of interest is incubated with a substrate peptide or protein, radiolabeled $[\gamma^{-32}P]ATP$ and MgCl.

Following separation of the unreacted $[\gamma^{-32}P]ATP$, the incorporation of ³²P into the substrate is quantified on a scintillation counter.

³²P incorporation assay

Detection of a radiolabeled phosphorylated substrate

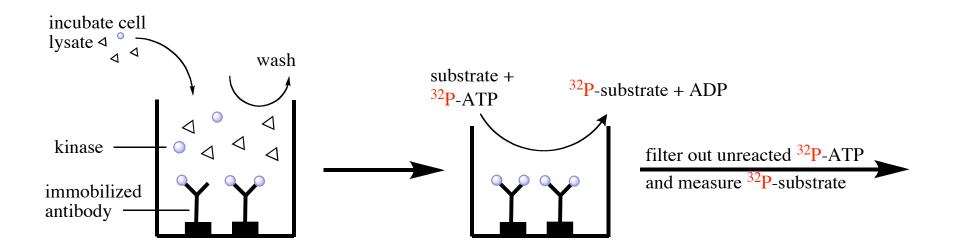


The requirement for residual ATP separation is the major drawback of this method. Additional assay considerations (both negative and positive):

- generates radioactive waste
- non-continuous, meaning that it is necessary to take time points if you need kinetic data.
- General and sensitive. The assay works for any kinase.

Note: ³²P incorporation (or other in-vitro) assays can be used for cell lysate if the kinase of interest is first purified from the cell lysate by immunoprecipitation.

Immunoprecipitation is the use of an antibody to specifically bind a target protein and provide a handle for protein isolation.



modified from Asthagiri, A. R. et al. *Anal. Biochem.* **269**, 342-347 (1999)

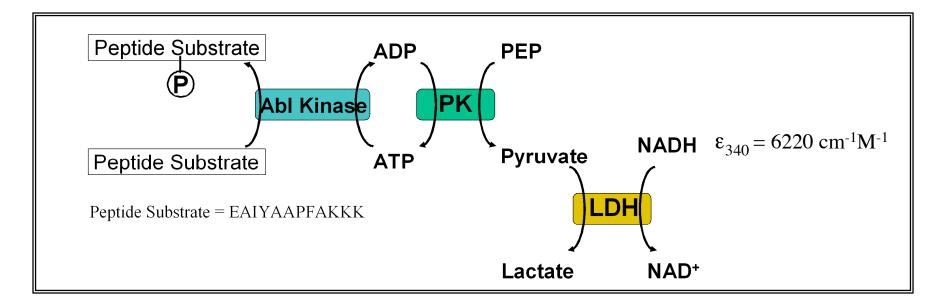
Gel Analysis with a Phospho-specific Antibody Western blot detection of a phosphorylated substrate

Ie. Detection of phosphorylated Ca concentrations of PDGF.	rk (an Abl substrate) from cells with increasing
5 5,	copyright restrictions. 7. et al. Genetically encoded fluorescent osine kinase activities in living cells. <i>PNAS</i> . 98
PDGF= platelet derived growth factor PDGF \rightarrow PDGFR \rightarrow Src \rightarrow Abl	Ting A. Y. et.al. <i>PNAS</i> 98 ,15003-15008 (2001)

Assay considerations (depending on your specific application)

- Non-continuous
- Often used qualitatively
- Highly specific and often used with crude cell lysate

Coupled Phosphorylation Assay (Session 13 and 14) (Indirect) detection of ADP formation/ATP consumption



Through two enzymatic reactions, Abl kinase activity is coupled to the conversion of NADH to NAD⁺.

The specific activity of the kinase can be calculated based on the decrease in NADH absorbance at 340 nm over time.

Extinction coefficient (*E*)

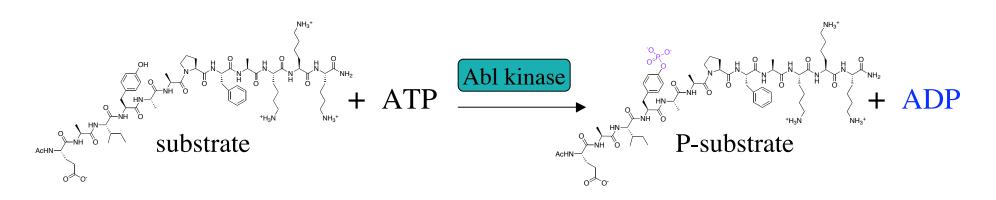
A molecule's extinction coefficient is a measure of the extent to which the molecule absorbs light at a given wavelength.

 ε is typically reported for the wavelength of maximum absorbance (ε_{max}) in units of cm⁻¹M⁻¹.

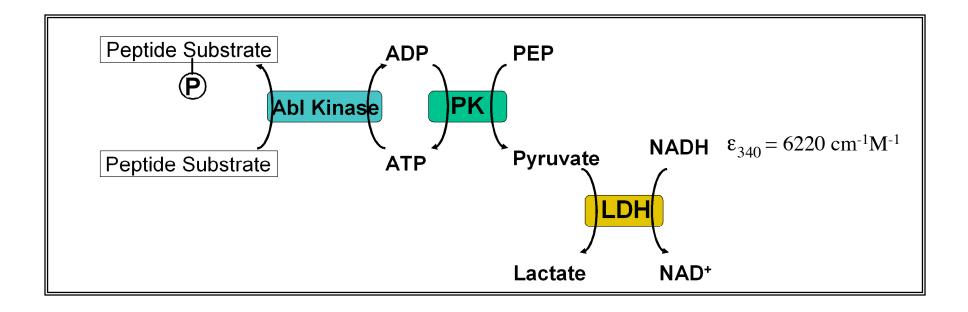
Beer's Law $Abs = \varepsilon cl$

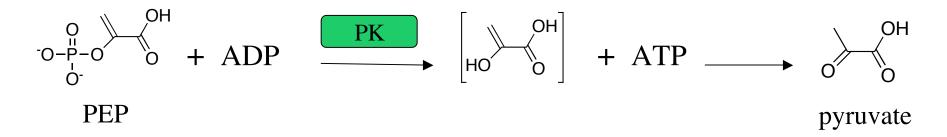
where c = concentration, and l = pathlength of the cuvette

Beer's law allows you to use ε to calculate the concentration of a sample in solution by measuring the Abs at a given wavelength.

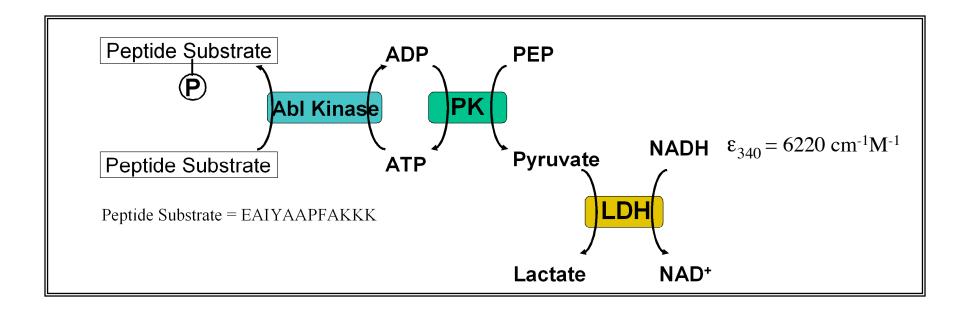


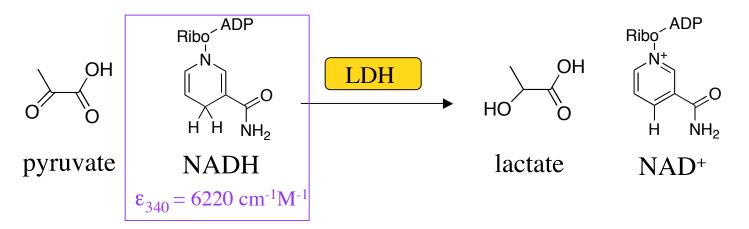
A protein kinase catalyzes the transfer of a phosphate group from ATP to a T, S, or Y on the peptide substrate. For our Abl assays, the substrate is $EAI\underline{Y}AAPFAKKK$.



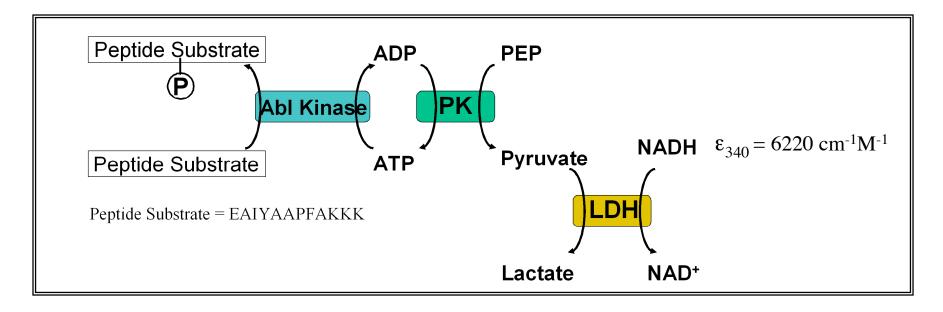


Pyruvate kinase (PK) transfers a phosphate group from phosphoenolpyruvate (PEP) to ADP to form pyruvate and ATP.





Lactate dehydrogenase (LDH) catalyzes the reduction of pyruvate to lactate with concomitant oxidation of the coenzyme NADH to NAD⁺.



You will monitor the loss of NADH absorption at 340 nm to quantify the specific activity of **wt** and **H396P** Abl kinase domain

- 1) in the absence of any inhibitors
- 2) with the Abl inhibitor Gleevec
- 3) with the Abl inhibitor Dasatinib

Specific activity in biochemistry is defined as the amount of product formed by an enzyme in a given amount of time.

Specific activity is often reported as:

units (U) per mg of enzyme,

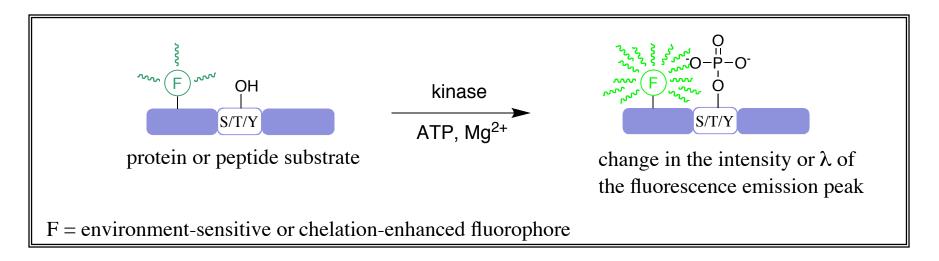
where 1 unit is equal to 1 μ mol of product formed per minute.

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Fluorescent Peptide-based Probes

Detection of the phosphorylated substrate

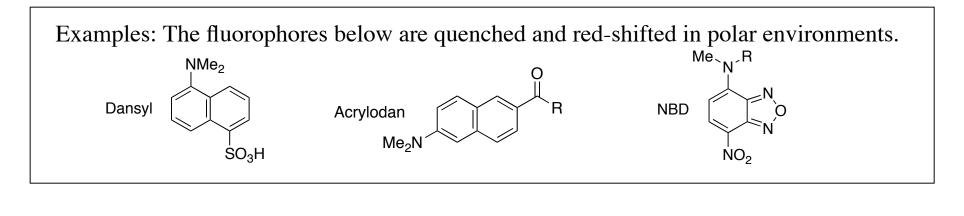


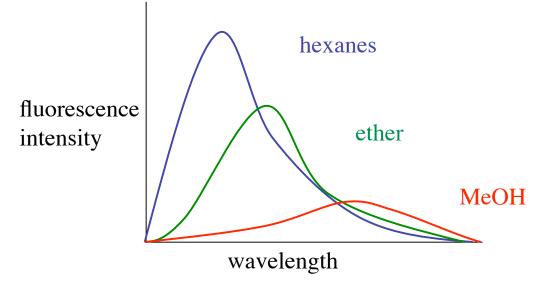
These probes comprise a substrate peptide that includes an amino acid recognition sequence for a specific kinase and an appropriately positioned environment-sensitive or chelation-enhanced fluorophore.

Upon phosphorylation of the substrate probe, the flurophore's emission properties change, enabling detection.

Fluorescent Peptide-based Probes: *Environment-Sensitive Fluorophores*

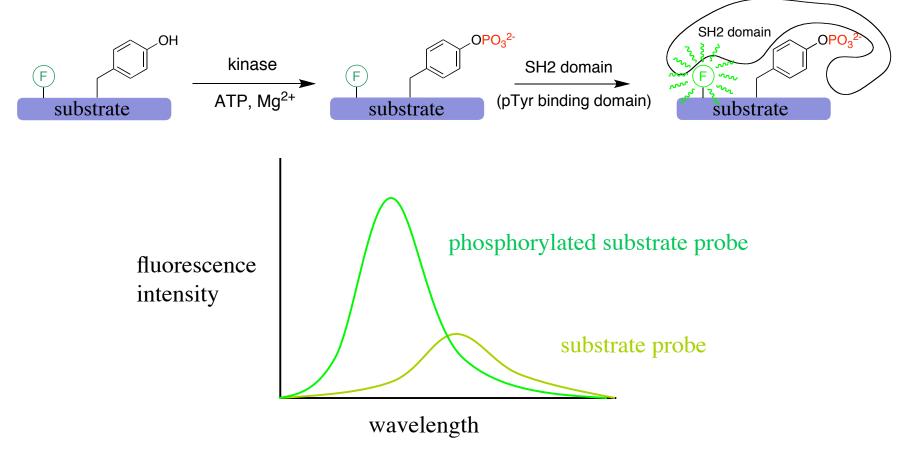
The signal intensity and maximum emission wavelength of environment-sensitive fluorophores (also referred to as solvatochromic dyes) are affected by the polarity of the fluorophore's immediate environment.





Fluorescent Peptide-based Probes: Environment-Sensitive Fluorophores

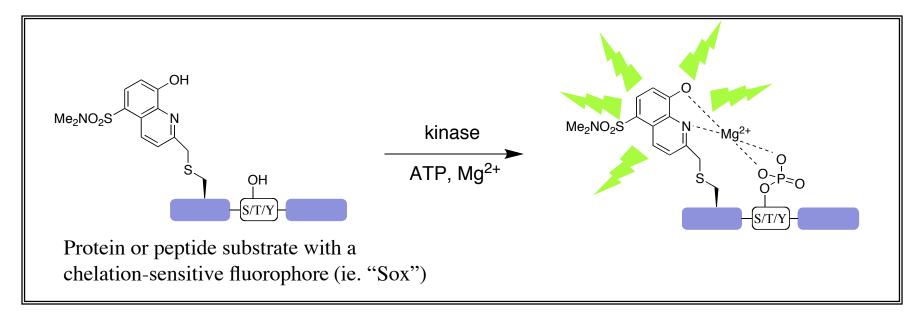
A phosphorylation-induced conformational change or intramolecular binding event that results in a change in the fluorophore's local environment can be detected by a change in fluorescence intensity.



Wang, Q. and Lawrence, D. S. J. Am. Chem. Soc. 1270, 7684-7685 (2005)

Fluorescent Peptide-based Probes: Chelation-Enhanced Fluorophores

Chelation-Enhanced Fluorophores demonstrate dramatic fluorescence intensity increases upon metal chelation.

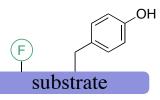


In CHEF-based kinase activity probes, the probes have low affinity for Mg²⁺ in the non-phosphorylated form. Once phosphorylated the binding affinity increases drastically, resulting in a 2 to 12 fold fluorescence enhancement.

Shults, M. D., Carrico-Moniz, D., and Imperiali, B. *Anal. Biochem.* **352**, 198-207 (2006) Lukovic, E., Gonzlez-Vera, J. A., and Imperiali, B. *J. Am. Chem. Soc.* **130**, 12821-12827 (2008)

Advantages to peptide based probes

- Assays enable continuous readings (no work-up steps prior to detection) making them ideal for kinetic studies.
- Peptide probes are amenable to large scale synthesis.
- Compared to proteins, peptides are incredibly robust for handling and storage.
- Analogs to sense different kinases and explore different sensing techniques are readily accessible.
- Fluorescence change upon phosphorylation in many peptide-based sensors greatly exceeds 100%.



Disadvantage

• It is challenging to uniformly introduce peptide probes into cells for in-vivo studies. (Most peptides are not cell-permeable.)

Introducing Peptide probes into cells (current methods) The major hurdle for using many of the peptide-based sensors in cell applications is introducing the sensor into the cell.

- microinjection
- Protein transduction domains (typically Arg-rich peptide sequences) that are fused to target peptides or small proteins and render them cell permeable.
- Introduction of a non-natural amino acid during protein expression (hijacking RNA translational machinery) for a genetically-encoded probe.

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Protein Probes

Protein probes for sensing kinase activity typically employ fluorescence resonance energy transfer (FRET) to sense a conformational change induced by protein phosphorylation.

FRET

FRET is a distance-dependent interaction between a donor and an acceptor fluorophore and is used as a "molecular ruler".

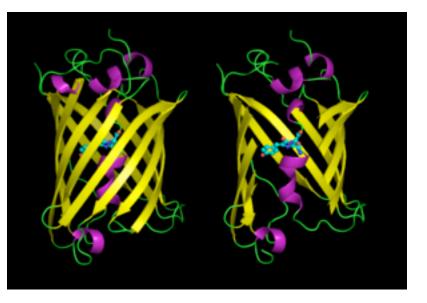
When the donor and acceptor fluorophores are in close proximity (10-100 A), the acceptor fluorophore is excited by the donor emission and emits a signal*.

fluorescence intensity shorter λ longer λ

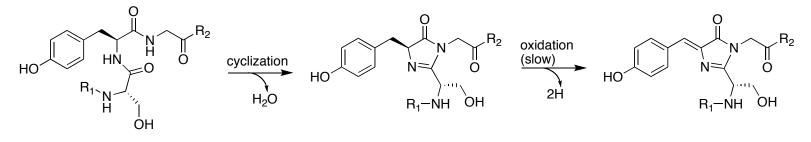
*In FRET, the emission spectrum of the donor must overlap with the absorption spectrum of the acceptor.

Green Fluorescent Protein (GFP) and related fluorescent proteins



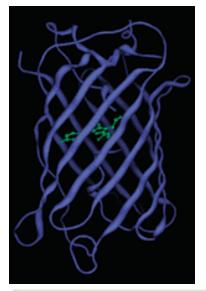


- 238 amino acid GFP first isolated from the Jellyfish Aequorea Victoria
- The chromophores of GFP and related fluorescent proteins arise from natural amino acids, so they can be expressed in-vivo.



<u>Ser</u>65 -<u>Tyr</u>66 - <u>Gly</u>67

Green Fluorescent Protein (GFP) and related fluorescent proteins



For (S65T) GFP: Excitation: 488 nm Emission: 507 nm

The denatured GFP protein is NOT fluorescent. The fluorophore is environment-sensitive.

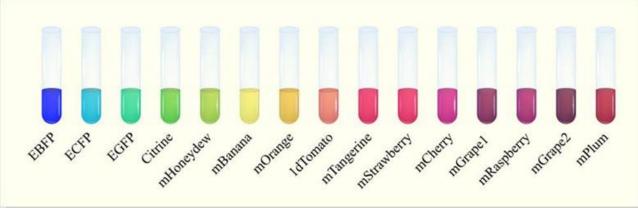


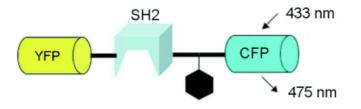
Image by MIT OpenCourseWare

Protein engineering has led to the development of a rainbow of other GFP analogs with a range of excitation and emission wavelengths. All are expressible, but also large (~230 amino acids).

FRET-based phosphorylation sensors:

Detection of a phosphorylation-induced conformational change

For example, below is a cartoon of a FRET-based sensor for protein tyrosine kinases (PTK) using the CFP/YFP FRET pair.

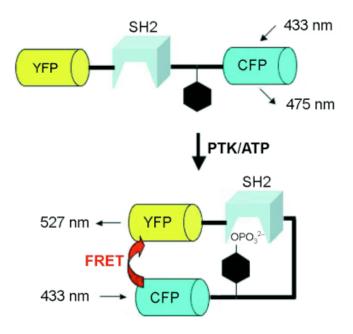


Lawrence, D. S. and Wang, Q. ChemBioChem 8, 373-378 (2007)

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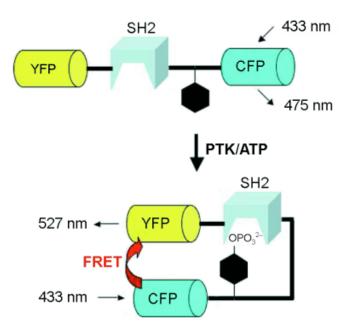
Lawrence, D. S. and Wang, Q. ChemBioChem 8, 373-378 (2007)

The major advantage to FRET-based probes is that they can be expressed in cells.

FRET-based phosphorylation sensors:

Detection of a phosphorylation-induced conformational change

For example, below is a cartoon of a FRET-based sensor for protein tyrosine kinases (PTK) using the CFP/YFP FRET pair.



However, the large size of FRET fluorophore pairs is a concern, particularly in comparison to small-molecule fluorophores.

Also, most FRET-based probes demonstrate a fluorescence increase of less than 60% following phosphorylation.

Example of a FRET sensor for Abl phosphorylation: The highest Abl activity is detected in PDGF-induced membrane ruffles

> Images removed due to copyright restrictions. See Fig. 3d and 3e in Ting, A. Y. et al. Genetically encoded fluorescent reporters of protein tyrosine kinase activities in living cells. *PNAS*. 98 (2001): 15003-15008.

PDGF= platelet derived growth factor PDGF \rightarrow PDGFR \rightarrow Src \rightarrow Abl

Ting A. Y. et.al. *PNAS* **98**, 15003-15008 (2001)

Laboratory Report Guidelines

Laboratory Report: **42 points** Abstract: **5 points** Introduction: **10 points** Explanation of purpose/relevance: (5) Demonstration of understanding: (5) Results: **5 points** Organization/Clarity: (5) Discussion: **10 points** Interpretation of data: (5) Demonstration of understanding: (5) Appendices: **12 points** PyMol Structure Worksheet: (4) Materials and Methods: (4) Supplementary Info: (4)

> Laboratory Work: **8 points** Overall technique: (4) Effort and attitude: (4)